



Droplet-on-Demand Platform for Biochemical Screening & Drug Discovery

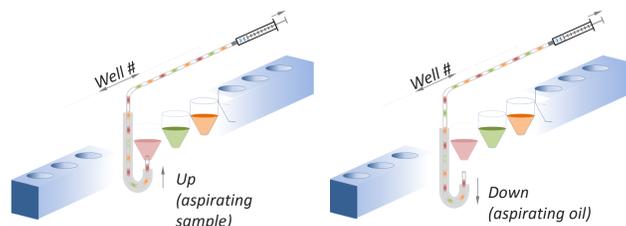
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Aim

To demonstrate droplet on demand applications towards study of biological entities encapsulated in nanoliter droplets. Interfacing a droplet on demand platform with microfluidic chips allows for merging and dilution of droplets. This feature is applied to encapsulate yeast cells (*S. cerevisiae*) and multi-cellular organisms (*C. elegans*).

Droplet on Demand Technology

Droplet Creation Mechanism



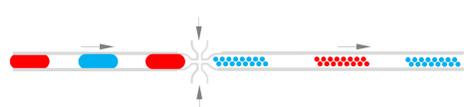
A constant aspiration is set up using a syringe pump. The sampling tube is programmed to move between aqueous and fluorocarbon fluids, thereby the creating plugs of the two immiscible fluids.

Droplet Merging and Splitting Mechanism

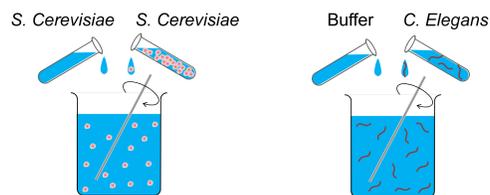
- Merging successive pairs after creating a sequence of nanoliter droplets



- Splitting each nanoliter droplet into picoliter droplets

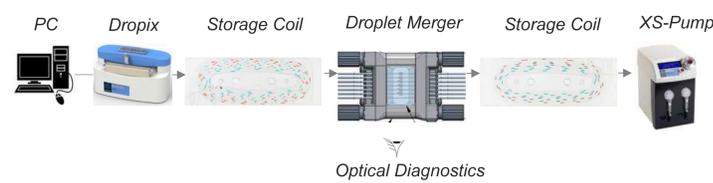


Method



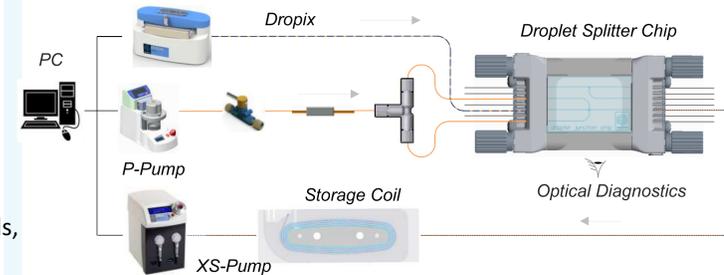
A culture of *S. Cerevisiae* cells is mixed in with cell buffer to make a resulting solution of 500 μ L. In a separate test, 5 day old *C. Elegans* embryos are mixed with buffer to make a dilute solution. 50 μ L of each is pipetted into the sample strip of the Dropix.

Test Setup 1 – Droplet Merging



- Setup consists of 1 syringe pump, 1 merging chip, 2 storage coils, and 1 Dropix. Syringe pump creates a suction driven flow.
- Flow rates range from 1 μ L/min to 20 μ L/min
- Merging chip has channel depth of 200 μ m
- Merged droplet volume ranges from 50 to 200 nL

Test Setup 2 – Droplet Splitting

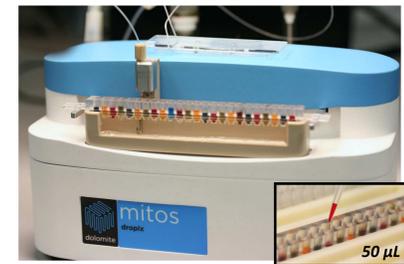


- Setup consists of 1 syringe pump, 1 droplet splitter chip, 2 storage coils, and 1 Dropix.
- Chip has a depth of 100 μ m. Flow rates 1 – 20 μ L/min.

Test Apparatus

Specifications

- Generates droplets of up to 24 different samples
- Droplet production frequency of 5Hz.
- Droplet size range from 50 nL to 50 μ L.
- Stores up to 1000 droplets.



MitoS Dropix – Droplet on demand platform. Each of the 24 fluid reservoirs holds up to 50 μ L.

The MitoS Dropix[®] system enables

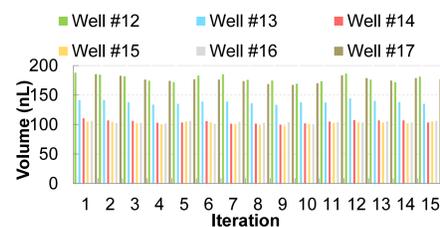
- Automated screening experiments (sample/screen pairing)
- Dose-response testing (sample/diluent pairing to deliver to screen)
- Concentration/Stoichiometric testing (variation of mix ratio)
- Combinatorial chemistry ($A_n B_n$ reagent pairings)

Results

Sequence Generation

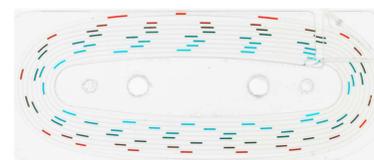


Droplet sequence of various sized nanoliter droplets.

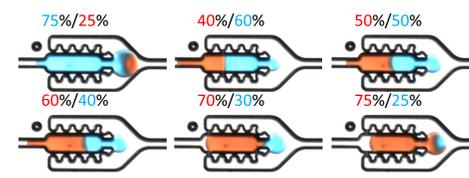


Multiple iterations of a 6 droplet sequence. Droplet volumes are 200, 150, 100, 100, 100, 200 nL.

Droplet Merging

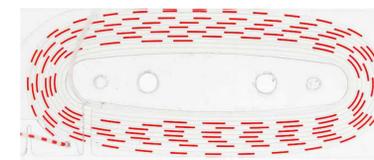


200 nL droplets. 100 step concentration gradient created from merging two liquids.

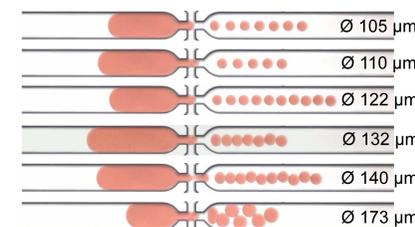


Red & blue droplets merging in varying proportions to create a concentration gradient.

Droplet Splitting



200 nL droplet sequence before splitting.



A nanoliter droplet split into multiple picoliter droplets.

C. Elegans Encapsulation



C. Elegans cells encapsulated in nanoliter droplets.

S. Cerevisiae Encapsulation



S. Cerevisiae cells encapsulated in nanoliter droplets.

References

[1] Gielen et al. A Fully Unsupervised Compartment-on-Demand Platform for Precise Nanoliter Assays of Time-Dependent Steady-State Enzyme Kinetics and Inhibition. *Anal. Chem.*, 85, 4761-4769, 2013.

[2] Niu et al. A microdroplet dilutor for high-throughput screening. *Nat. Chem.*, 3, 437-442, 2011.

[3] Bilsland et al. Yeast-based automated high-throughput screens to identify anti-parasitic lead compounds. *Open Biol.*, 3, 120158, 2013.

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Future Outlook

- Merging yeast cells with different drugs has the potential to show different growth patterns of pooled yeast strains.
- This will be a valuable tool for drug selectivity studies [3]. *C. Elegans* can be encapsulated using our system, and merging encapsulated organisms with drugs could be used to study effects on neural mechanism.

Acknowledgements

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